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Molecular Markers

What are Molecular Markers?

- Specific fragments of DNA that can be identified within the whole genome. Molecular markers are the general assays that allow detection of the sequence differences between two or more individual. Molecular markers are found at specific locations of the genome. They are used to 'flag' the position of a particular gene or the inheritance of a particular characteristic or desired characteristics
- molecular marker is a DNA sequence that is readily detected and whose inheritance can easily be monitored .

Molecular Taxonomy

- RAPD (Random Amplified Polymorphic DNA)
- RFLP (restriction fragment length polymorphism)
- ISSR (Inter-simple sequence repeat)
- SSR (simple sequence repeat)
- ITS (Internal transcribed spacer)

- DNA markers Non-PCR Based, RFLP- Restriction fragment length polymorphism. PCR Based RAPD- Random amplification of polymorphic DNA . AFLP -Amplified fragment length polymorphism. SCAR -Sequence characterize amplified region. STS - Sequence tagged sites. EST- Express sequence tags. SNP- Single nucleotide polymorphism. SSR -Simple sequence repeats, ISSR, ITS CAPS -Cleaved amplified polymorphic sequences etc.

- Shortly after Kary Mullis invented the Polymerase Chain Reaction (PCR) it was realized that short primers would bind to several locations in a genome and thus could produce multiple fragments
- Williams *et al.* (1990) developed Random Amplified Polymorphic DNA (RAPD) a technique using very short 10 base primers to generate random fragments from template DNAs

- Template DNA Primers point in the same direction, so amplification won't happen
- Template DNA Primers too far apart, so amplification won't happen > 2,000 bases
- Template DNA Primers are just the right distance apart, so fragment is amplified 100 - 1,500 bases

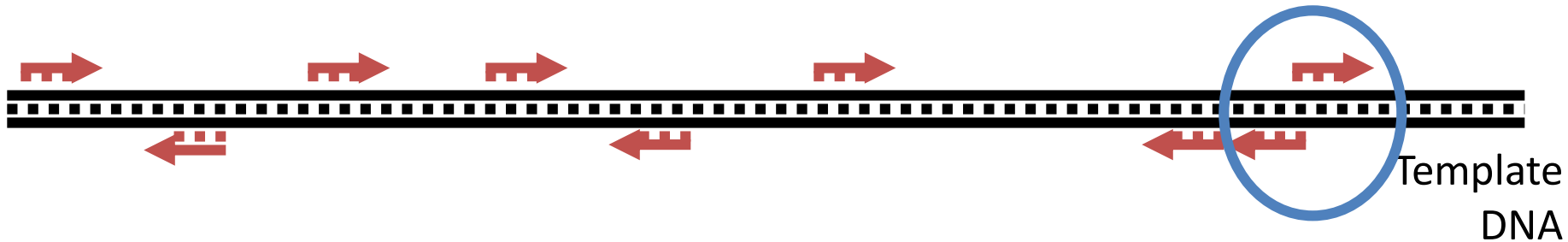
- RAPD fragments can be separated and used as genetic markers or a kind of DNA fingerprint
- RAPD polymorphism result from change in the primer-binding site in the DNA sequence
- In variety A there are 4 primer binding sites resulting in two RAPD products, variety B lacks one of the binding sites resulting in only one RAPD marker being produced

1. Buffer (containing Mg^{++}) - usually high Mg^{++} concentrations are used lowering annealing stringency
2. Template DNA
3. 1 short primer (10 bases) not known to anneal to any specific part of the template DNA (2 Primers that flank the fragment of DNA to be amplified as in the case of PCR)
4. dNTPs
5. *Taq* DNA Polymerase (or another thermally stable DNA polymerase)

Modifying Thermal Cycling

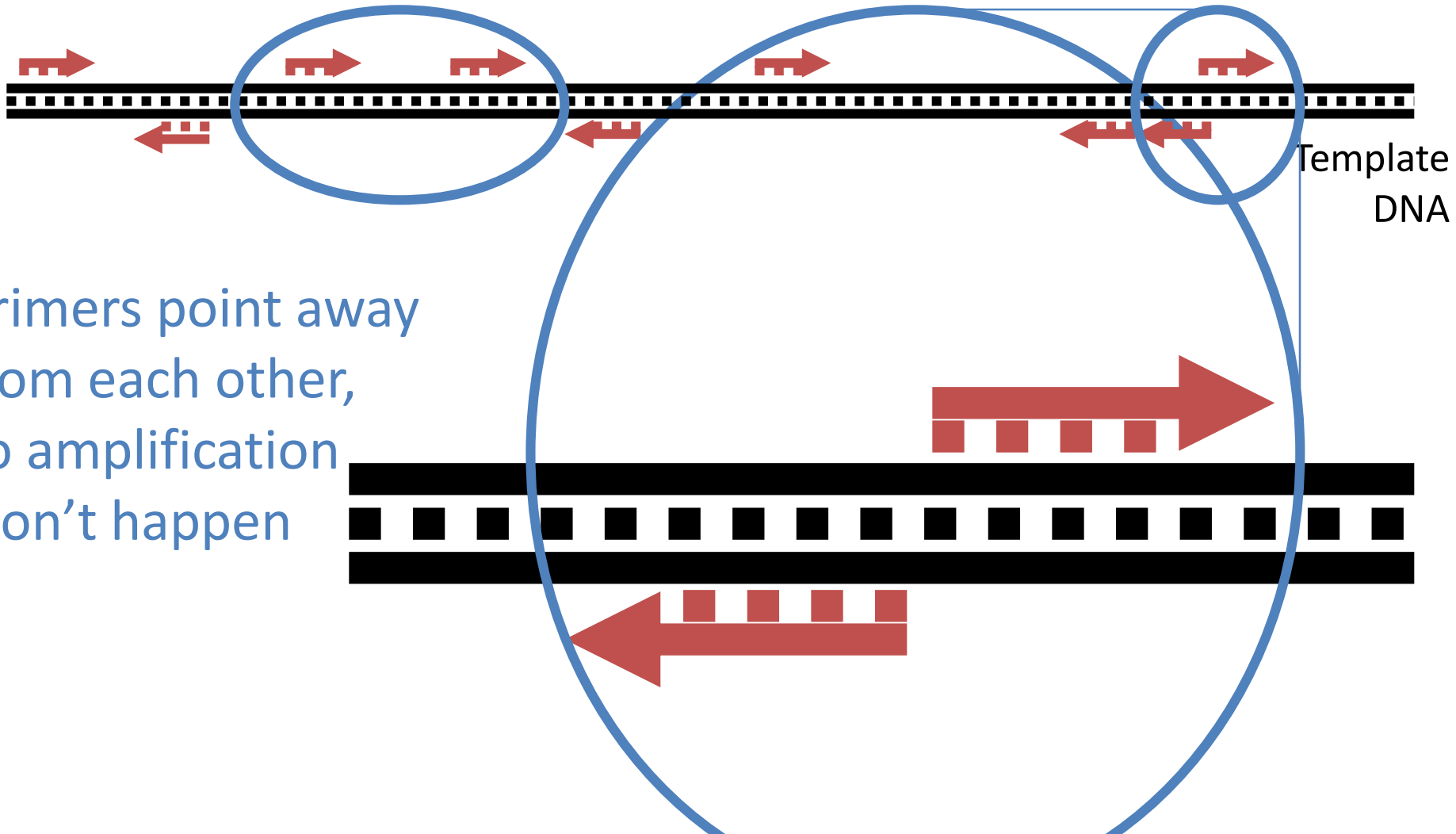
- Two modifications made to typical thermal cycling when RAPD is being done:
 1. Annealing temperatures are generally very low, around 36 °C - This allows very short primers to anneal to template DNA
 2. More thermal cycles are used, typically 45 - This compensates for the inefficiency which results from using such short primers.

RAPD



- Primer binds to many locations on the template DNA
- Only when primer binding sites are close and oriented in opposite direction so the primers point toward each other will amplification take place

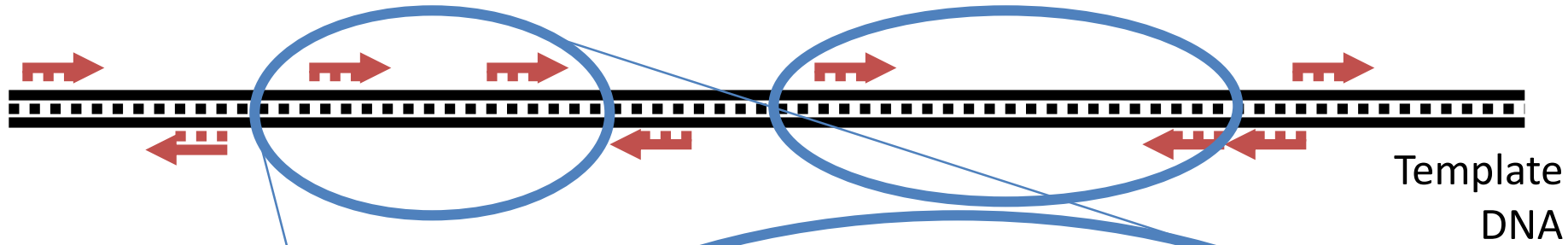
RAPD



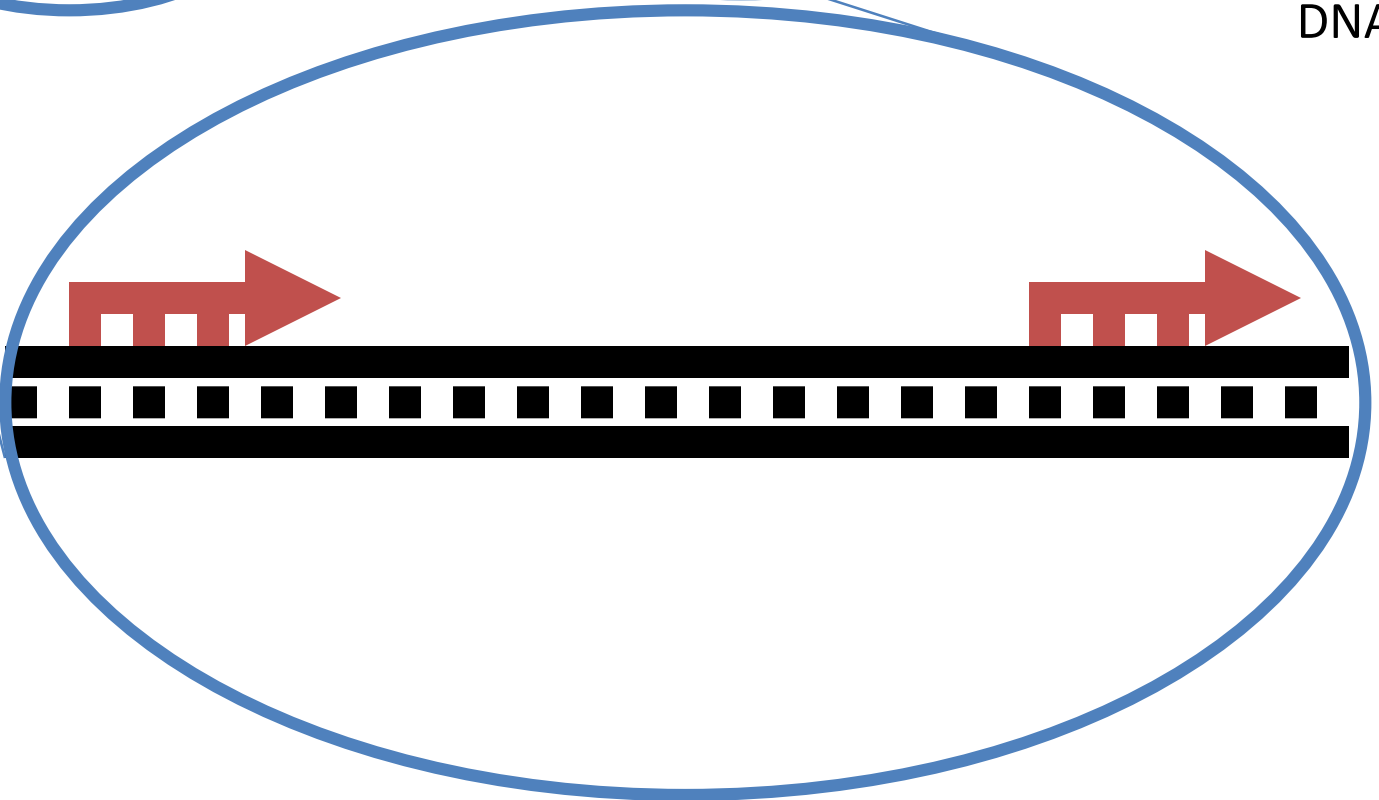
Template
DNA

Primers point away
from each other,
so amplification
won't happen

RAPD

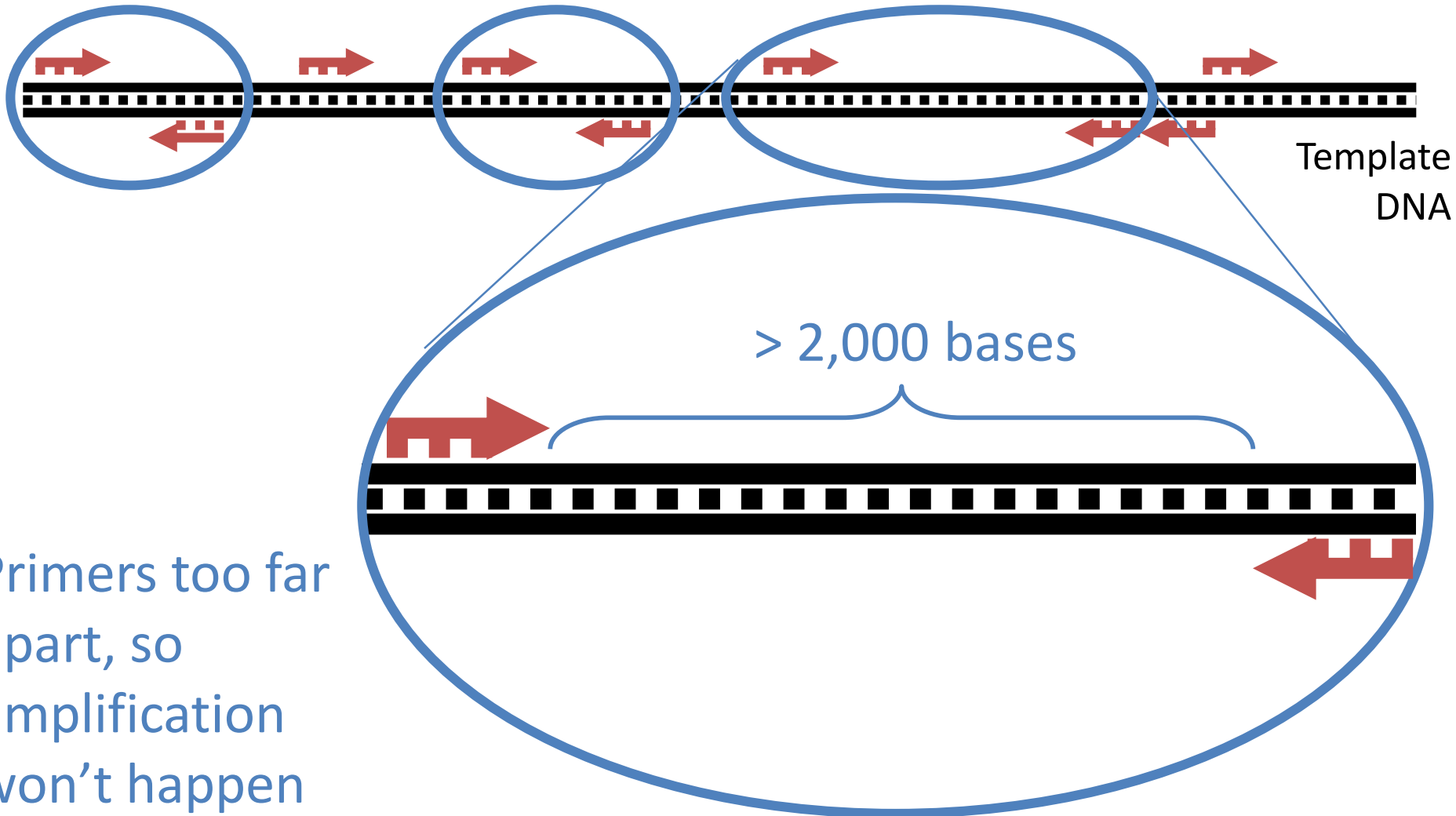


Template
DNA



Primers point in
the same
direction, so
amplification

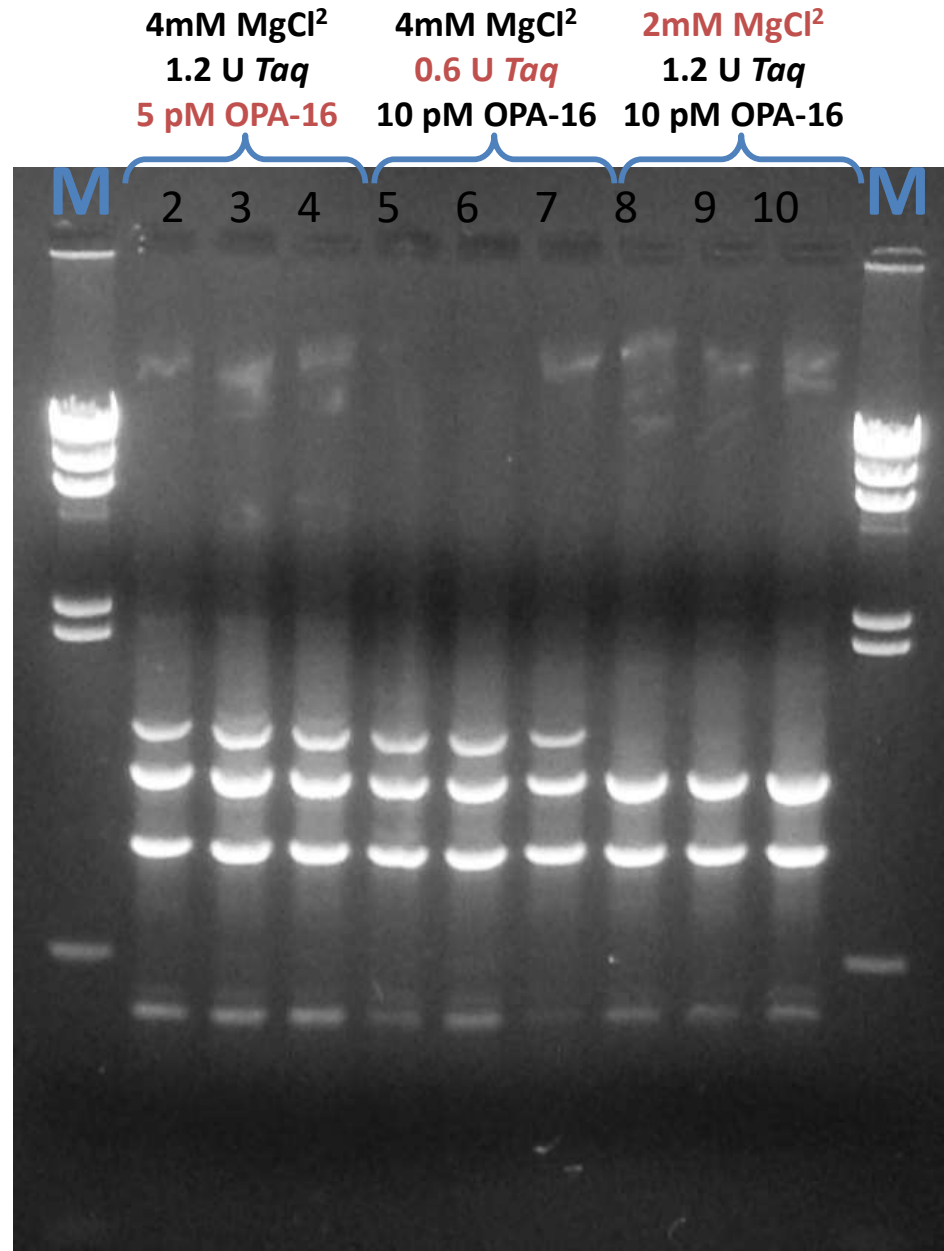
RAPD



Separated RAPD Fragments

RAPD reactions were run in groups of 3 using the same template and primer, but varying Magnesium, polymerase and primer concentrations

Normal concentrations are shown in yellow text. M = A size standard

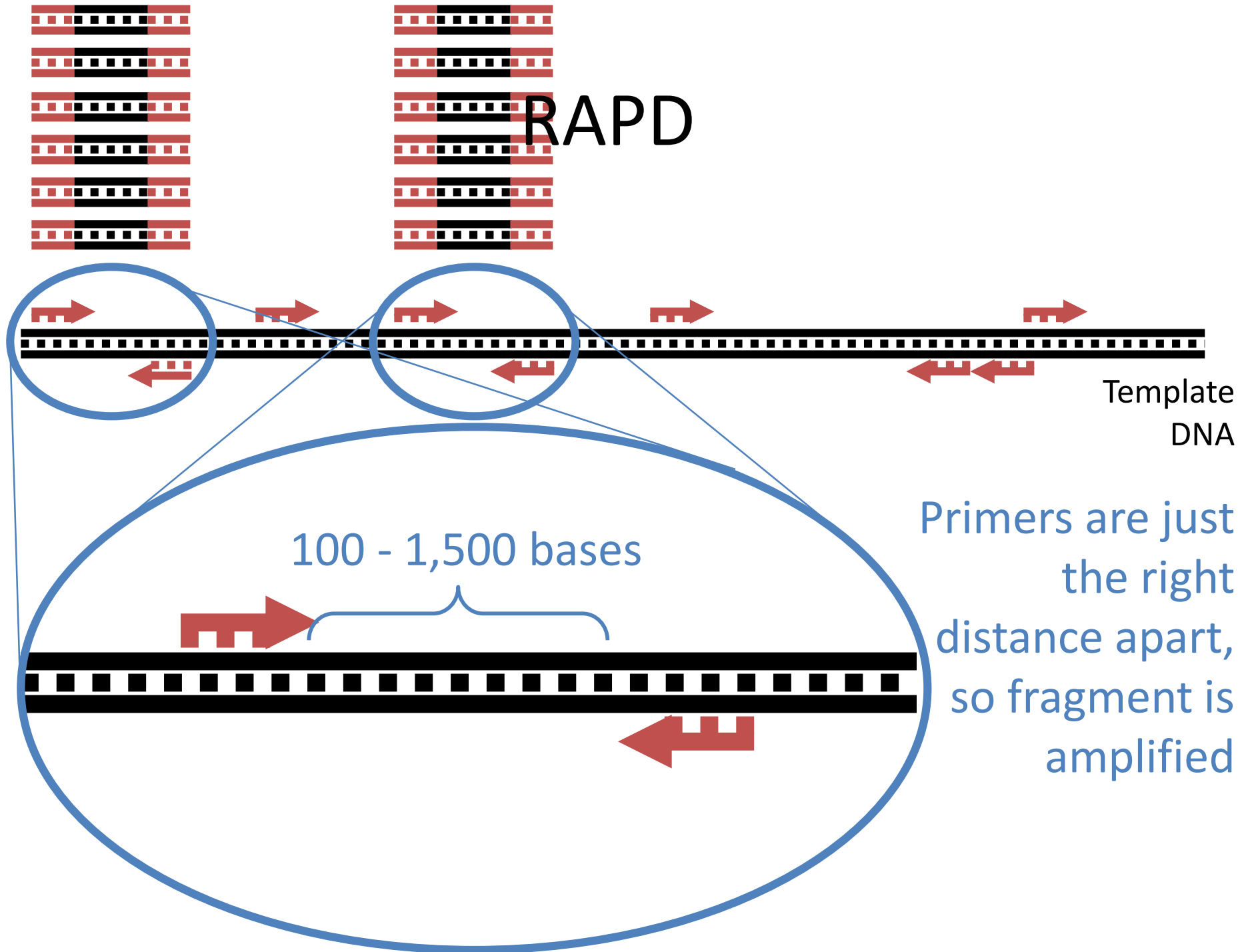


Which variable has the greatest impact on fragment patterns?

Lowering Magnesium ion concentration results in loss of the largest fragment visible in lanes 2-7

need small amount of DNA -it involves non-radioactive assay -it does not required specific probe libraries -it provide quick and efficient screening for DNA sequence based on polymorphism at many loci -it is inherited as dominant traits -there is a bands due to relatively short primer -the production of non-parental bands in the offspring of known pedigree warrants its use with extreme care -it is sensitive to change in PCR conditions
Advantages Disadvantages

RAPD



Template DNA

100 - 1,500 bases

Primers are just the right distance apart, so fragment is amplified

RFLP

- The variation(s) in the length of DNA fragments produced by a specific restriction endonuclease from genomic DNA s of two or more individuals of a species RFLP

- RFLP technology was first developed in the 1980s for use in human genetic applications and was later applied to plants. By digesting total DNA with specific restriction enzymes, an unlimited number of RFLPs can be generated. RFLPs are relatively small in size and are co-dominant in nature. If two individuals differ by as little as a single nucleotide in the restriction site, the restriction enzyme will cut the DNA of one but not the other. Restriction fragments of different lengths are thus generated. All RFLP markers are analyzed using a common technique. However, the analysis requires a relatively complex technique that is time consuming and expensive.

- The hybridization results can be visualized by Autoradiography (if the probes are radioactively labeled), or Chemiluminescence (if non-radioactive, enzyme-link methods are used for probe labeling and detection). Any of the visualization techniques will give the same results.

- simple and easy to use -easy to detect via PCR
-co-dominant marker -perfectly suited for
used in map-based cloning -cost is higher for
establishing polymorphic primer sites and
investment in the synthesizing the
oligonucleotides -initial identification, DNA
sequence information necessary Advantages
Disadvantages

- Application of SSR Assessment of genetic variability and characterization of germplasm. Identification and fingerprinting of genotypes. Estimation of genetic distances between population, inbreds and breeding material. Marker assisted selection. Identification of sequence of useful candidate genes

SSR

- Microsatellites can be amplified for identification by the polymerase chain reaction (PCR) process, using the unique sequences of flanking regions as primers. DNA is repeatedly denatured at a high temperature to separate the double strand, then cooled to allow annealing of primers and the extension of nucleotide sequences through the microsatellite. This process results in production of enough DNA to be visible on agarose or polyacrylamide gels. With the abundance of PCR technology, primers that flank microsatellite loci are simple and quick to use, but the development of correctly functioning primers is often a tedious and costly process. Principle

Thank You