Core course BMS361N Genetic Engineering

## In vitro mutagenesis

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### In vitro mutagenesis

- Many methods are available
  - Random mutagenesis
  - Site directed mutagenesis
  - Deletion mutagenesis
  - Insertion mutagenesis

 Random mutagenesis is used generally to screen particular genetic functional analysis

### Mutagenesis

# Mutagenesis

- One can clone gene to express it in a specific host organism to produce a large amount and very pure protein that can be used commercially.
- However physico chemical properties of these proteins (which are naturally occurring) are often not suited for particular task at extreme environment.
- Ex. Glucose oxidase, purified naturally occurring enzyme's optimal temperature is at 37oC but if one commercialize this, harsh handling, changing environment may inactivate. To over come this type of problem, molecular biologist came up with idea to create or modify or changing aa encodes an enzyme with desired properties so that which can be used at both extreme conditions or particular task.

By using a set of techniques what specifically change as encoded by a cloned gene Protein with properties that are better suited than naturally occurring enzymes. Ex.

By altering both substrate binding and maximal rate of conversion of the substrate to product.

By changing the thermal tolerance or pH stability or both.

By changing an enzyme so that a cofactor is no longer required for large scale reaction.

By changing substrate binding site to increase its specificity, so that non-specific reactions and its products are reduced.

By increasing resistance to cellular protease, so that protease cannot act on it thereby increase the yield of proteins.

By altering the allosteric regulation to diminish the impact of feedback inhibition and increase the yield.

#### **Mutants**

#### Deletions

 One can delete one nucleotide to few hundred nucleotides depending upon their experimental need.

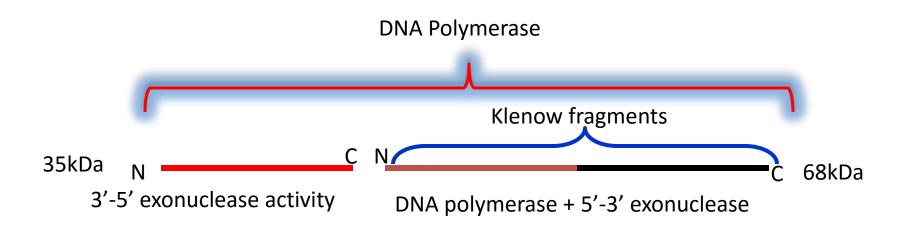
#### Insertions

 One can insert one nucleotide to few hundred nucleotide depending upon their experimental need.

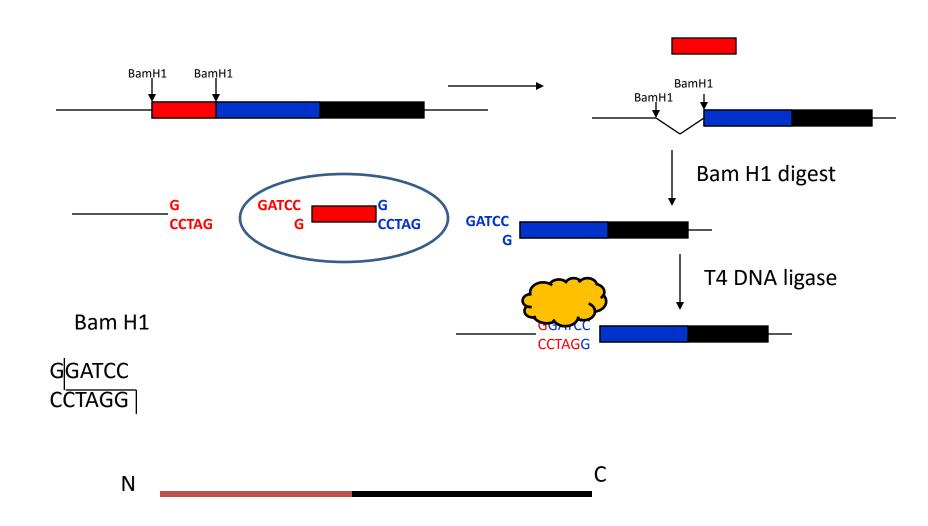
#### Deletion

### Deletion

 If RE site present in site where one wants to make deletion, they can use RE to digest and remove part of the DNA fragment from the cloned DNA fragments

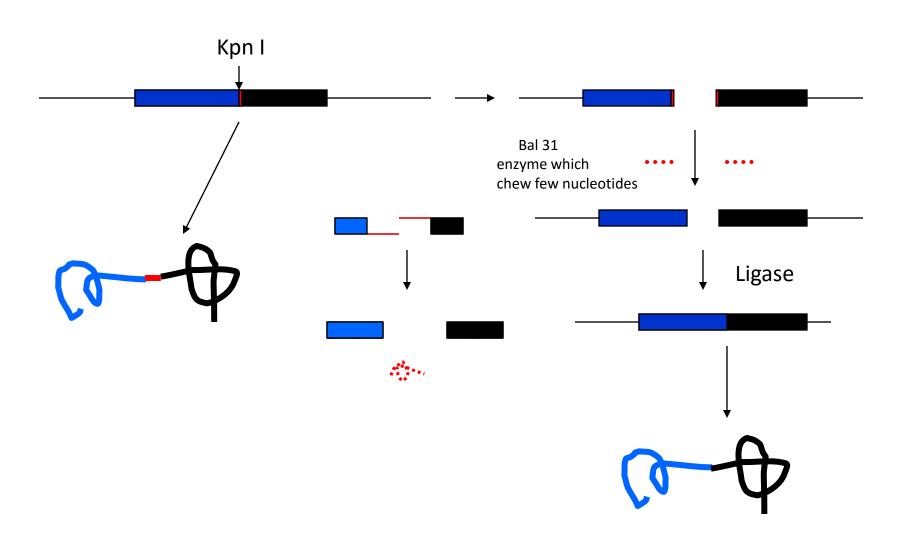


### Few nucleotides Deletion

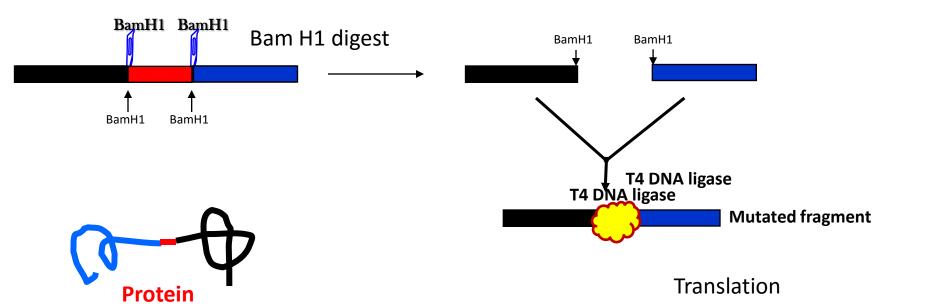


#### Few nucleotides Deletion

# Removing few nucleotide



### Deletion



Bam H1

G GATCC CCTAG G **Mutated Protein** 

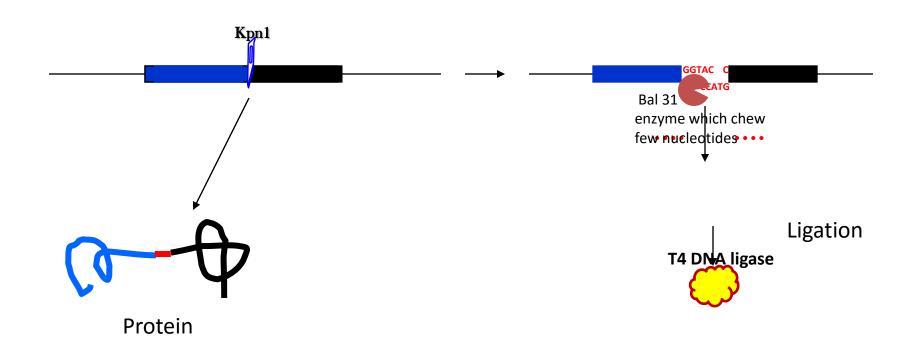
#### Few nucleotides Deletion



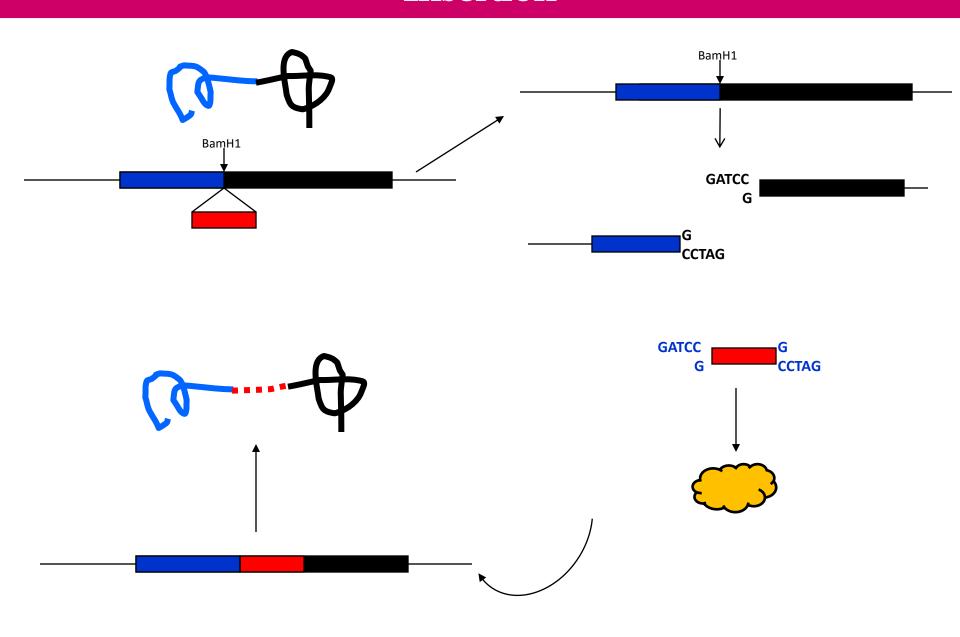




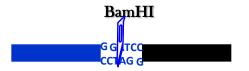
# Removing few nucleotide



### Insertion



### Insertion





### Site directed mutagenesis

- By many method one can achieve this mutagenesis.
- Main mechanism is Oligonucleotides can be synthesized and used to make mutant DNA.
   This mutant DNA used to express mutant protein and then analyzed the mutant induced effect by comparing wild type protein.

### Site directed mutagenesis

## Site Directed mutagenesis

- Site directed mutagenesis:
- A single nucleotide (point mutation) change can make various functions
- Ex. Single nucleotide can make
  - » non-sense mutant
  - » Sense mutant
  - » Frame shift

## The Good, the Bad, and the Silent

Mutations can serve the organism in three ways:

A mutation can cause a trait that enhances the organism's function:

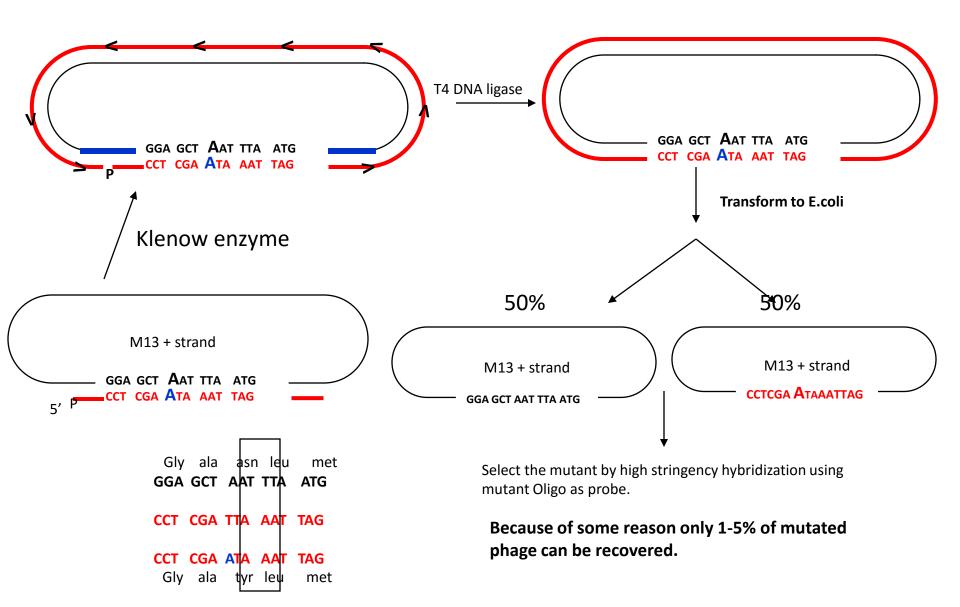
- The Good: Mutation in the sickle cell gene provides resistance to malaria.
- The Bad:

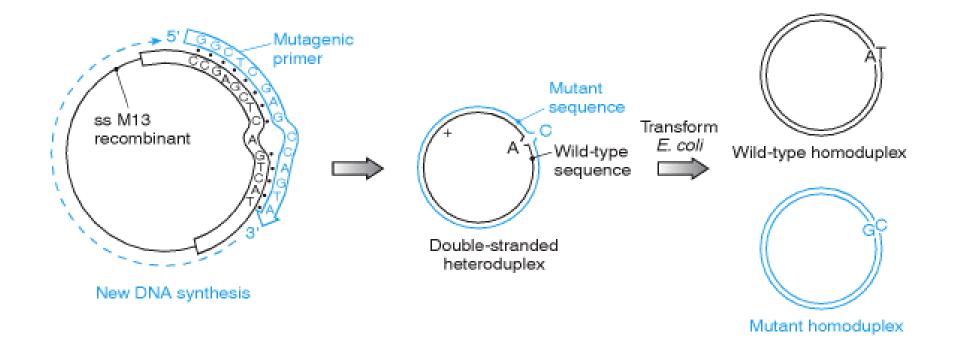
  A mutation can cause a trait that is harmful, sometimes fatal to the organism:

Huntington's disease, a symptom of a gene mutation, is a degenerative disease of the nervous system.

• The Silent: A mutation can simply cause no difference in the function of the organism.

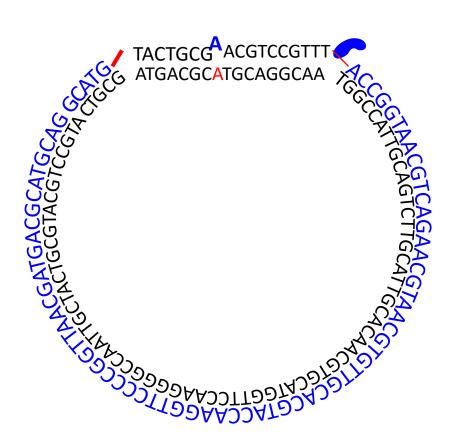
#### **Plasmid based**

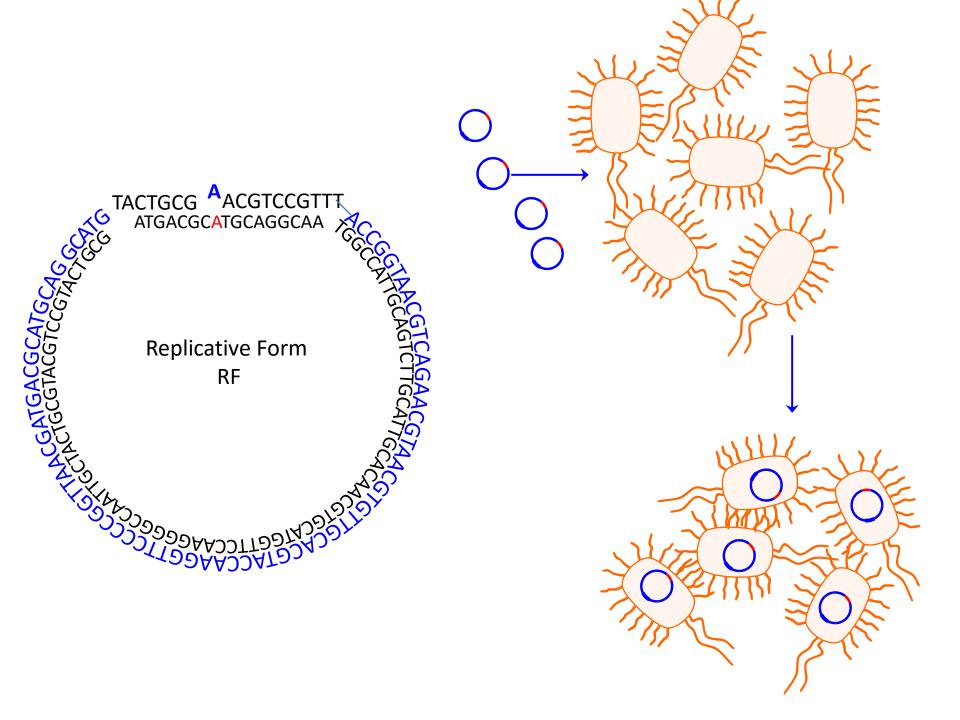


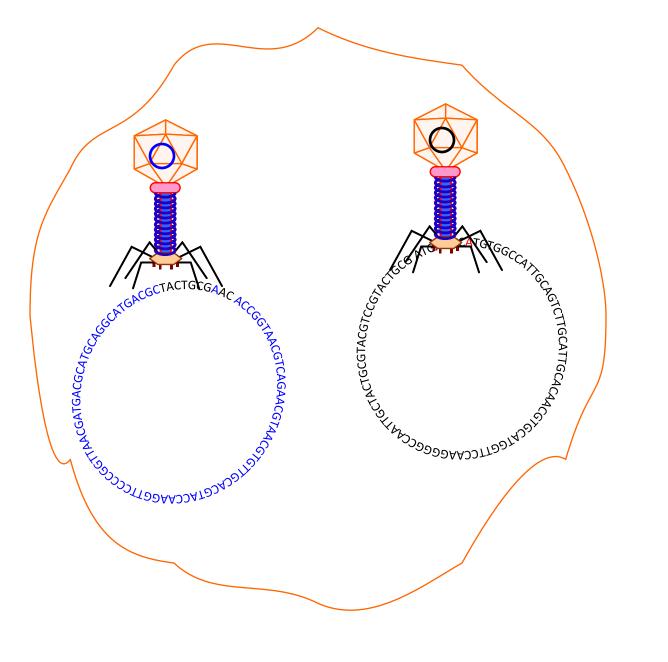


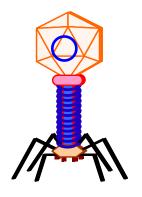
Site Directed mutagenesis

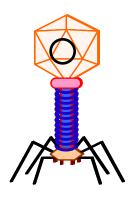


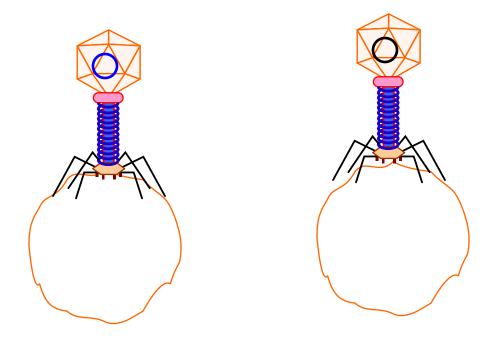












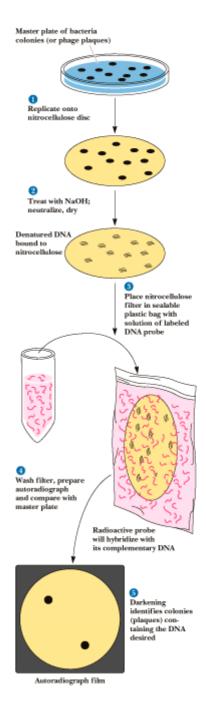
By hybridization method one can isolate mutated Plasmid.

## Colony Hybridization

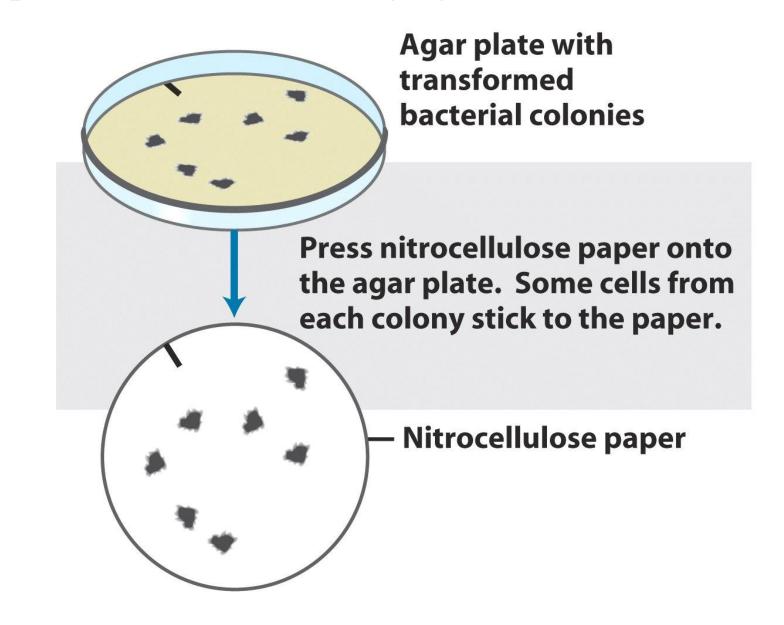
A way to screen plasmid-based genome libraries for a DNA fragment of interest

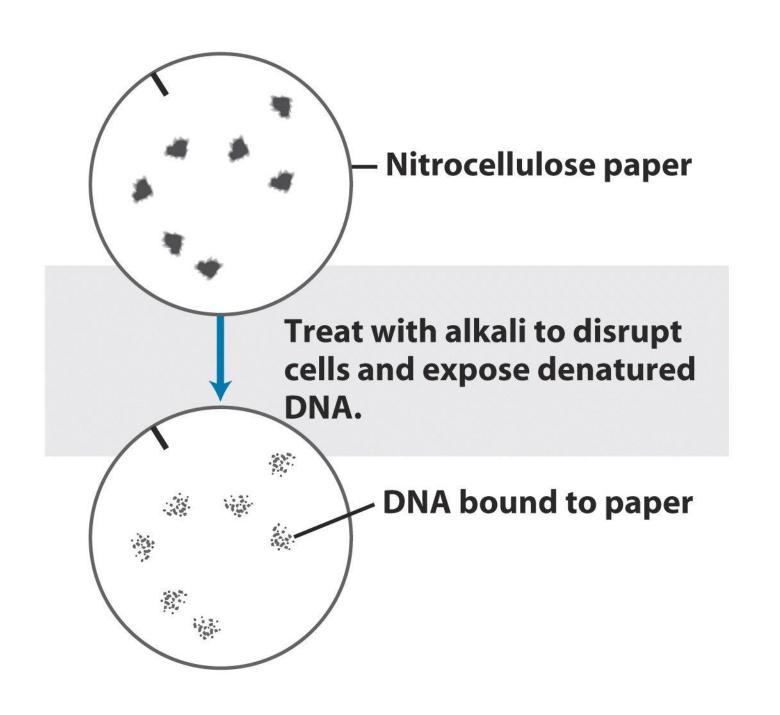
- Host bacteria containing a plasmid-based library of DNA fragments are plated on a petri dish and allowed to grow overnight to form colonies
- Replica of dish made with a nitrocellulose disk

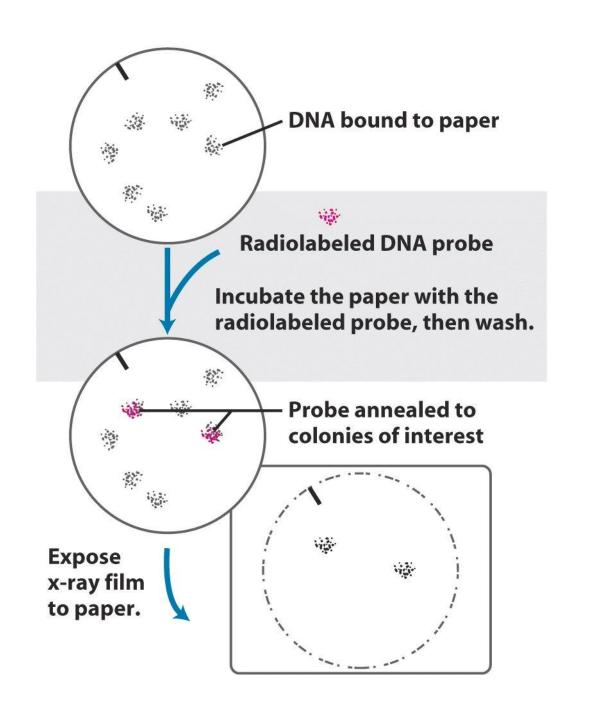
- Disk is treated with base or heated to convert dsDNA to ssDNA and incubated with probes
- Colonies that bind probe (with P-32) hold the fragment of interest



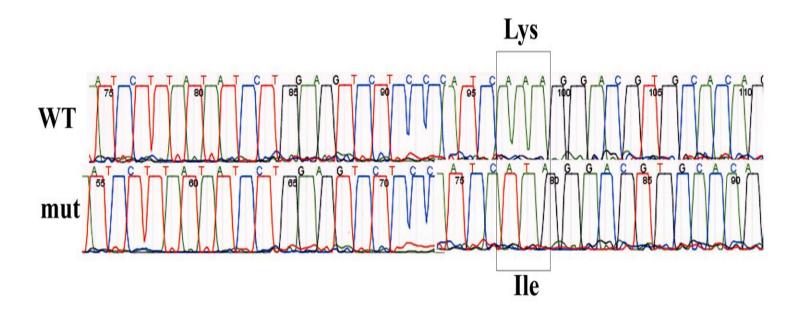
#### Specific sequences are detectable by hybridization







# Confirmation by DNA sequencing



## Modified Oligo-directed mutageneis

Oligo-directed mutagenesis is modified in a number of ways to enrich number of mutant phage.

One of the method is:

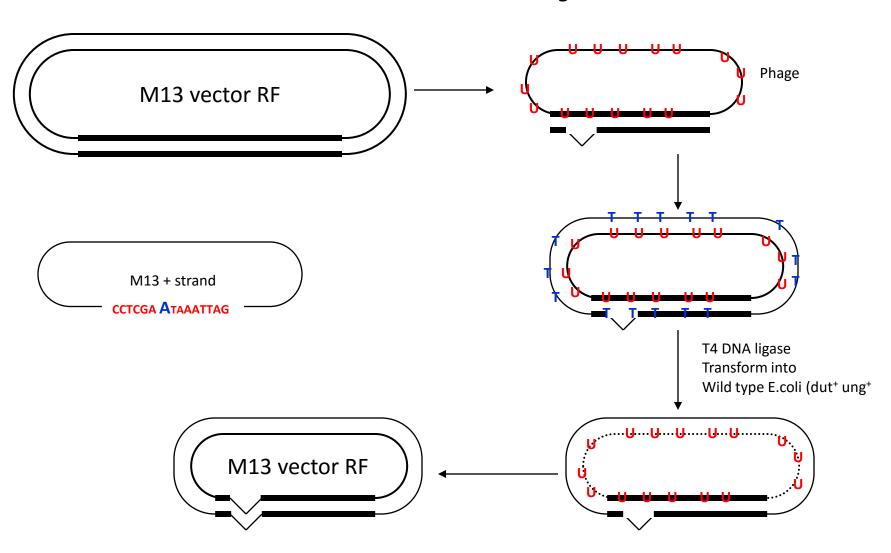
Use of different E.coli strains one of the E.coli strain has 2 defective DNA metabolism enzymes

- (1) defective form of dUTPase (*dut*⁻)
- (2) defective form of Uracil N-glycosylase (ung<sup>-</sup>)

Because of these defective enzymes incorporation of UTP residue cannot be remove. So the genomic DNA and vectors transformed into this E.coli strains have UTPs in the place of TTPs. By make use of these properties one can enrich mutant vectors.

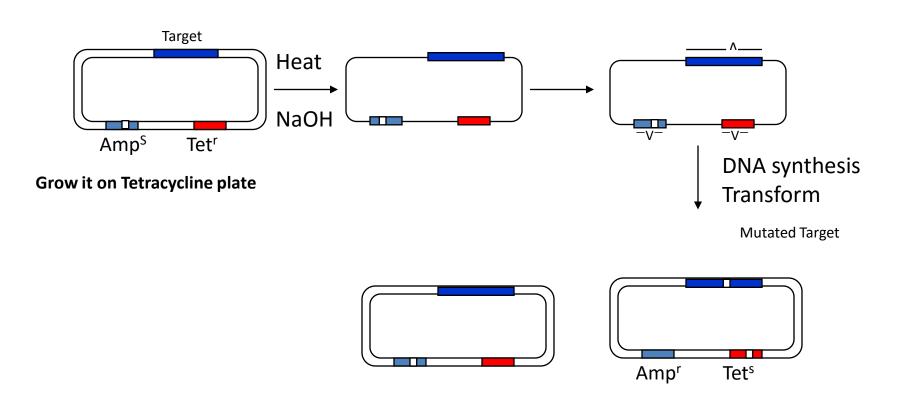
## Modified Oligo-directed mutageneis

Transform to E.coli **dut** - **ung** -



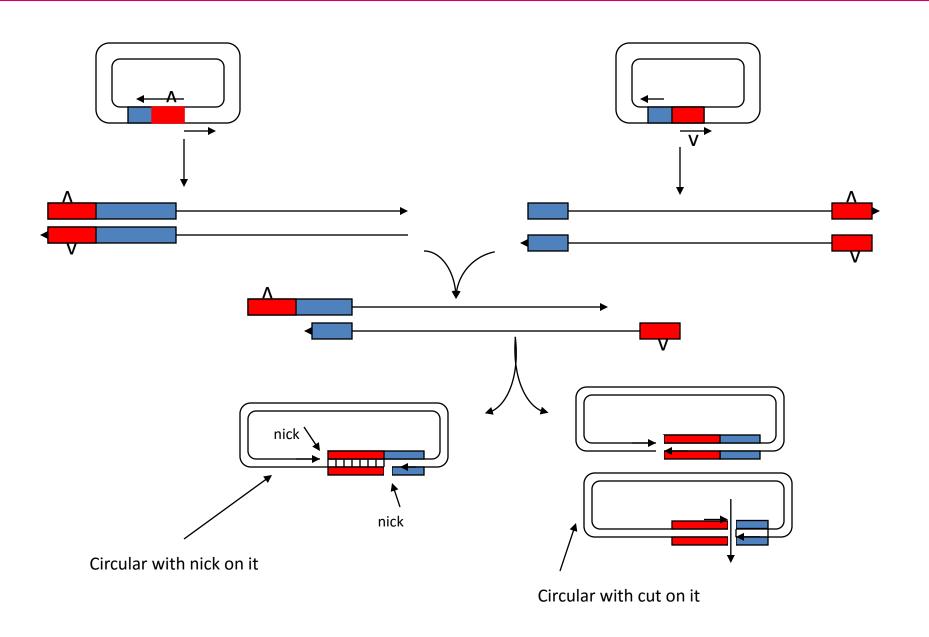
## Modified Oligo-directed mutageneis

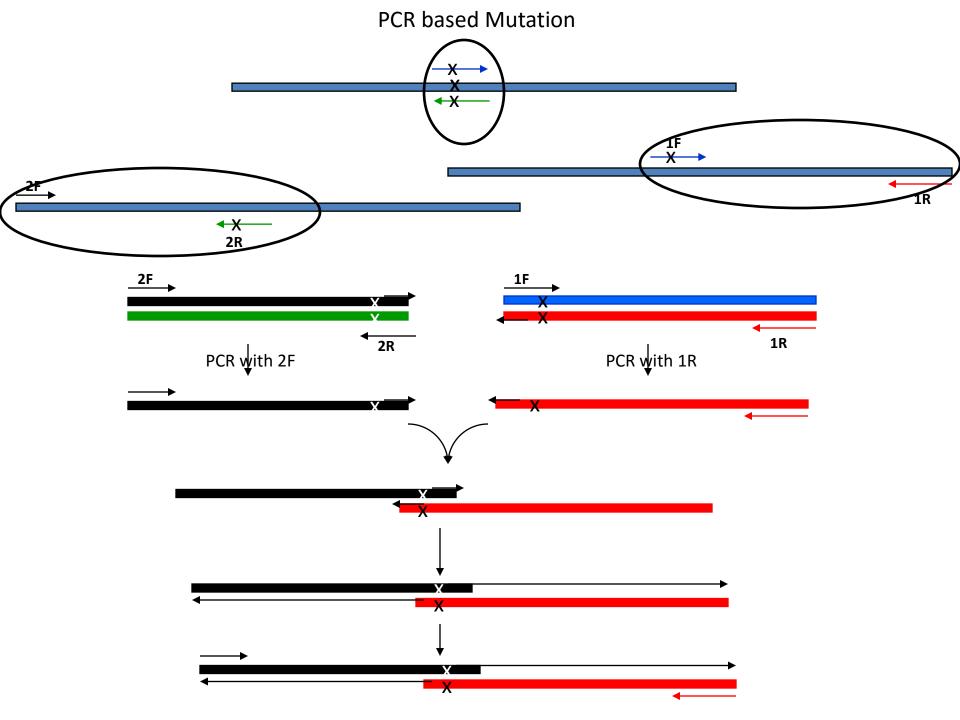
Using Ds Plasmid



Grow it on ampicilin plate

### PCR based Oligonucleotide directed mutagenesis





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#### The End

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